

The TGF- β Signaling Pathway as a Pharmacological Target in Hepatocellular Carcinoma

Antonio Mazzocca, Salvatore Antonaci and Gianluigi Giannelli*

Department of Emergency and Organ Transplantation, Section of Internal Medicine, Allergology and Clinical Immunology, University of Bari School of Medicine, 70124 Bari, Italy

Abstract: Hepatocellular carcinoma (HCC) is a cancer that usually develops on a liver already compromised by cirrhosis. Study of the underlying molecular mechanisms is essential so as to improve therapeutic strategies and to develop new pharmacological agents that may prevent or improve the course of this malignancy. Transforming growth factor-beta (TGF- β) intervenes in the process of hepatic fibrogenesis and cirrhosis, two pathogenic preconditions for the formation and progression of HCC [1] [2]. In addition, TGF- β plays a crucial role in the molecular pathogenesis of HCC and may, therefore, prove to be a promising drug target. We and other authors have recently demonstrated that inhibition of the TGF- β signaling pathway results in a synergistic downstream action with an inhibitory effect on the progression of HCC. Several TGF- β inhibitors have recently been developed, most of which are still in a preclinical phase, but they may soon be available for testing in patients with HCC. However, well-designed clinical trials will be needed to evaluate the effectiveness of these new agents prior to routine use in the clinic. Aim of this article is to make a brief review of the benefits and limitations of targeting the TGF- β signaling pathway in HCC.

Keywords: Hepatocellular carcinoma, inhibitors of the TGF- β kinase receptor, mechanism-based drugs, stroma-tumor interactions.

1. INTRODUCTION

Research into new mechanism-based drug therapies for the treatment of patients with hepatocellular carcinoma (HCC) has become more competitive since sorafenib was introduced for the treatment for HCC patients with advanced disease [3]. As in other malignancies, the issue is to identify suitable pharmacological molecular targets and agents that exert little or no cytotoxicity. In HCC, this is particularly desirable in view of the presence of the liver cirrhosis which is commonly associated to HCC and limits the use of many drugs. Even the use of a multi-target inhibitor, like sorafenib, that fights the cancer via inhibiting several signaling pathways, can be limited by numerous side effects. In fact, this type of drug should, in theory, display more side effect compared to drugs directed against only a single target. Tailoring the therapy to the individual patient would be a promising strategy to circumvent this problem [4]. For this reason, many efforts have been made to achieve molecular characterization of those patients who are eligible for treatment with a certain drug. In theory, this could be achieved by identifying the specific gene signature, or even better a kinases signature (i.e. a profile of activity of protein kinases involved in response to the drug). An alternative strategy would be to drive preclinical research faster in the direction of identifying the mechanisms responsible for HCC development and progression. This could lead to the identification of key molecules actively participating in the process of tumor progression, that could then be targeted by appropriate drugs [5]. Clearly, to achieve results that can be translated to humans, such preclinical research should provide reliable *in vivo* models of HCC which replicate, at least in part, the tumorigenic phases occurring in humans. Although animal models of liver cirrhosis and hepatitis viruses are still lacking, a wider range of animal models is now available to study HCC [6,7]. These animal models are becoming useful tools for pharmacological studies in HCC and will hopefully improve the preclinical findings to be translated to clinical studies.

2. PATHOGENESIS OF HCC

Fibrotic liver disease and chronic inflammation often precede the onset of HCC by about 10 or 20 years, which suggests that these may act as a precancerous condition for HCC. Therefore, HCC can be seen as a multi-step carcinogenetic process [2]. The role of chronic viral hepatitis (HBV and HCV) in HCC is well-established [8]. These pathological conditions are characterized by varying degrees of fibrogenesis, considered as a predisposing factor for the onset of HCC [9]. As in other tissues, fibrosis of the liver is considered to be partly a physiological response triggered to repair liver damage. However, when the liver fibrosis becomes chronic, the repair mechanisms and inflammation associated with this chronic situation can become irreversible and lead to the condition of cirrhosis. Some pathological aspects of liver cirrhosis are very important, such as nodular hyperplasia, which is considered a premalignant lesion from which HCC develops [10]. Typically, HCC is a multinodular disease even when it is first detected as a single node. The majority of HCC forms are well-circumscribed nodular tumors, round or oval, without a capsule. Four distinct microscopic patterns have been identified: (1) fibrolamellar; (2) pseudoglandular (3) pleomorphic/giant cell, (4) clear cell. HCC presents more differentiated hepatocyte trabeculae, cords and nests, and contains pigments or bile, while poorly differentiated tumors show malignant cells that are not cohesive, pleomorphic, or anaplastic giant cells. Because of these important features, the histology and tumor staging will take into account the size and differentiation of the lesions, and will include the assessment of serum α -fetoprotein, the spread (for example, the presence of vascular invasion and the presence of intra- and extrahepatic metastases) and the degree of tumor vascularization (angiogenesis). These aspects are important to estimate the expected survival and determine the type of intervention. In an attempt to combine the clinical and laboratory predictors of outcome in patients with HCC, several staging systems have recently been proposed, including Barcelona Clinic Liver Cancer staging (BCLC) that is currently the most widely approved and recommended for use in clinical trials [11,12].

3. ROLE OF TGF- β 1 IN TUMOR SUPPRESSION AND CANCER PROGRESSION

Clinical data suggest that TGF- β 1 may play an important role in HCC progression. This is not the only consideration that could jus-

*Address correspondence to this author at the Department of Emergency and Organ Transplantation, Section of Internal Medicine, Allergology and Clinical Immunology, University of Bari School of Medicine, Bari, Italy; Tel: ++ 39 080 5478-127; Fax: ++ 39 080 5478-126; E-mail: g.giannelli@intmed.uniba.it

tify the use of an inhibitor of the TGF- β 1 pathway in the treatment of HCC. In fact, TGF- β 1 is involved in the process of fibrogenesis and tissue remodelling of the microenvironment where HCC develops [13]. Indeed, from a cancer biology standpoint, TGF- β 1 is an intriguing molecule because of its dual role as a tumor promoting and tumor suppressor factor. In fact, it can promote the metastatic dissemination of cancer cells in the late phases of tumor progression while, as a tumor suppressor, it can exert an anti-proliferative effect during the early phase of epithelial tumorigenesis [14,15,16]. The role of TGF- β 1 as a tumor suppressor has been widely described [17,18,19]. It is noteworthy that TGF- β 1 has been shown to be a pleiotropic factor with multi-functional activities [20,21,22]. Because of its role in regulating tissue homeostasis in HCC and because of its essential role in cancer formation, inhibition of the TGF- β 1 pathway remains a promising therapeutic strategy in this disease. A summary of the multifunctional is shown in (Fig. 1) activity of TGF- β 1 during HCC progression. Ideally, blocking the TGF- β 1-mediated pathway would reduce the fibrogenesis and prevent both the development of liver cirrhosis and the formation of HCC. The interaction between tumor and stroma is fundamental for the development of the tumor burden. In the case of HCC, this is clearly evident since liver cirrhosis is the most important risk factor. For example, it has been shown that connective tissue growth factor (CTGF), a protein regulated by TGF- β 1, acts as a fundamental regulator of fibrogenesis leading to the accumulation of extracellular matrix proteins in the liver and kidney [23] [24]. In cancer, CTGF has been reported to promote tumor growth, angiogenesis, migration and invasion [25]. Consistently with this observation, our research group has recently demonstrated that TGF- β 1-induced CTGF leads to the formation of highly stromogenic tumors in a human xenograft model of HCC. On the contrary, the same tumors show a poor stromal component when CTGF is knocked down by RNA interference [26]. Most importantly, the presence of abundant stroma affects the clinical outcome of these tumors, in the sense that the stromogenic HCC were bigger, grew faster and invaded more efficiently. As expected, inhibition of the TGF- β 1 pathway with LY2109761 inhibited CTGF production and blocked the growth of HCC. Histologically, these tumors show a less marked stromal component. In this scenario, the interaction between tumor and the surrounding tissue microenvironment, by means of cancer-associated fibroblasts (CAF), that produce high levels of CTGF in response to paracrine signals from cancer cells, may be a molecular target for therapeutic strategies [26]. In accordance with our findings, the pro-tumorigenic role of Cancer-associated fibroblasts (CAF) has recently been reported in different malignancies [27,28]; and more recently in HCC [29]. For example, Tumors with abundant CAF are resistant to chemotherapy, likely because the extracellular matrix components could contribute to cancer cell survival [30] [31].

4. ROLE OF TGF- β 1 IN FIBROGENIC LIVER DISEASES AND HCC

The role of TGF- β in liver fibrogenic disease and HCC is well-documented. The TGF- β signaling pathway consists of three distinct ligands, TGF- β 1, TGF- β 2, and TGF- β 3. All three ligands can bind to a specific receptor, firstly engaging with TGF- β RI, which then heterodimerizes with TGF- β RII. This heterodimer complex phosphorylates the intracellular Smad2 and 3 proteins, which in turn gives rise to a cascade of activation to induce different nuclear proteins transduction [32]. TGF- β has been investigated because of its association with organ fibrosis and collagen synthesis, where it is evaluated as a potential marker [33,34]. In fact, TGF- β 1 levels are elevated in fibrosis associated with both viral and alcoholic hepatitis. Moreover, in patients with documented HBV serum levels, TGF- β 1 levels are associated with fibrosis, and in HCV patients who respond to treatment with interferon-alpha and ribavirin, the TGF- β 1 levels decrease [35,36,37].

Liver fibrosis consists of an accumulation of stiff, fibrous scar tissue in the liver. This process is associated to chronic viral or alcoholic hepatitis and becomes more prominent in liver cirrhosis. Hepatic stellate cells (HSC) are specialized pericytes characterized by their ability to store retinol, that play a central role in liver fibrogenesis [38] [39]. After acute or chronic injury affecting hepatocytes, HSC are activated and transdifferentiate into myofibroblasts [40]. As a consequence, these cells activate an autocrine or paracrine TGF- β loop. Upon TGF- β stimulation, HSC produce extracellular matrix (ECM), and in particular proteins such as fibronectin and collagen [41] [42]. Moreover, the secretion of TGF- β creates a response by the immune system that prevents the down-regulation of the inflammatory process in the liver [43] [44] [45,46]. Gene expression and functional studies show that in patients with chronic liver fibrosis, liver tissue with a rich TGF- β content contains a large number of regulatory T cells [47,43,44].

As pointed out above, TGF- β has a dual role, acting as a tumor suppressor that regulates cell proliferation, but also as a tumor promoter facilitating the motility and invasion of cancer [48]. This dual effect is well known but is still controversial, and how and why TGF- β acts differently in the different stages of cancer progression is still unknown. One of the first studies on TGF- β 1 levels in HCC was performed in six patients who showed an increased TGF- β expression at mRNA and protein level [49]. A few years later, the same authors were able to correlate the tissue expression with increased plasma levels of TGF- β 1 in patients with HCC, and these levels have been shown to be higher than in patients with chronic hepatitis and cirrhosis [50]. In addition, the levels of TGF- β 1 decreased in patients treated for liver cancer, suggesting its potential use as a marker for monitoring the response of patients to treatment. In addition to plasma levels, urinary TGF- β 1 levels were also found to have prognostic value, resulting in poor survival in patients with HCC [51]. Moreover, high levels of TGF- β 1 were found in patients with aggressive and metastatic HCC as compared with patients with non-metastatic HCC [52]. Because a high vascular density is linked to poor survival, it is not surprising that high tumor vascular density is associated with high levels of TGF- β 1, and then correlated with aggressive tumor growth [53] [54]. Patients with vascular invasion display worst prognosis and shorter survival [55,56,57,58]. Measurement of TGF- β 1 levels in plasma of patients with HCC is feasible and may have a prognostic value [59] [51]. To increase the prognostic value of TGF- β 1 levels, it is possible to combine the levels of TGF- β 1 with other protein markers. When TGF- β 1 is combined with α -fetoprotein (AFP) it seems to improve the diagnostic predictability [60,61]. However, the difficulty of obtaining a reproducible prognostic marker is not only linked to the heterogeneous nature of the cancer, but also to the fact that sample recovery and analysis must be performed with great care and precision, to avoid confusing the liver-derived TGF- β with the platelet-derived TGF- β isoform, for example. As in liver fibrosis, TGF- β levels increases in HCC, along with collagen deposition and the reduction of proteolytic degradation [62]. Another factor with stromogenic properties, modulated by TGF- β in the liver, is connective tissue growth factor (CTGF) [63]. CTGF promotes tumor growth, angiogenesis, migration and invasion [64] [25]. HCC human cell lines produce high levels of CTGF to form highly stromogenic tumors. If CTGF is knocked down in these cells, tumors show a poor stromal component [26]. Blocking the TGF- β signaling pathway with the TGF- β inhibitor LY2109761 inhibits CTGF production and tumor growth. Based on these observations, cancer-associated fibroblasts (CAF) are considered a possible source of CTGF in response to paracrine signals from tumor cells, such as TGF- β [64,65,66].

5. INHIBITION OF TGF- β SIGNALING IN HCC

Pharmacological inhibition of TGF- β 1 has recently been proposed by different authors as an anti-cancer therapeutic approach [67,68];[69], Clin Cancer Res 2007; [70]. We have proposed that pharmacological inhibition of TGF β R can be considered a new

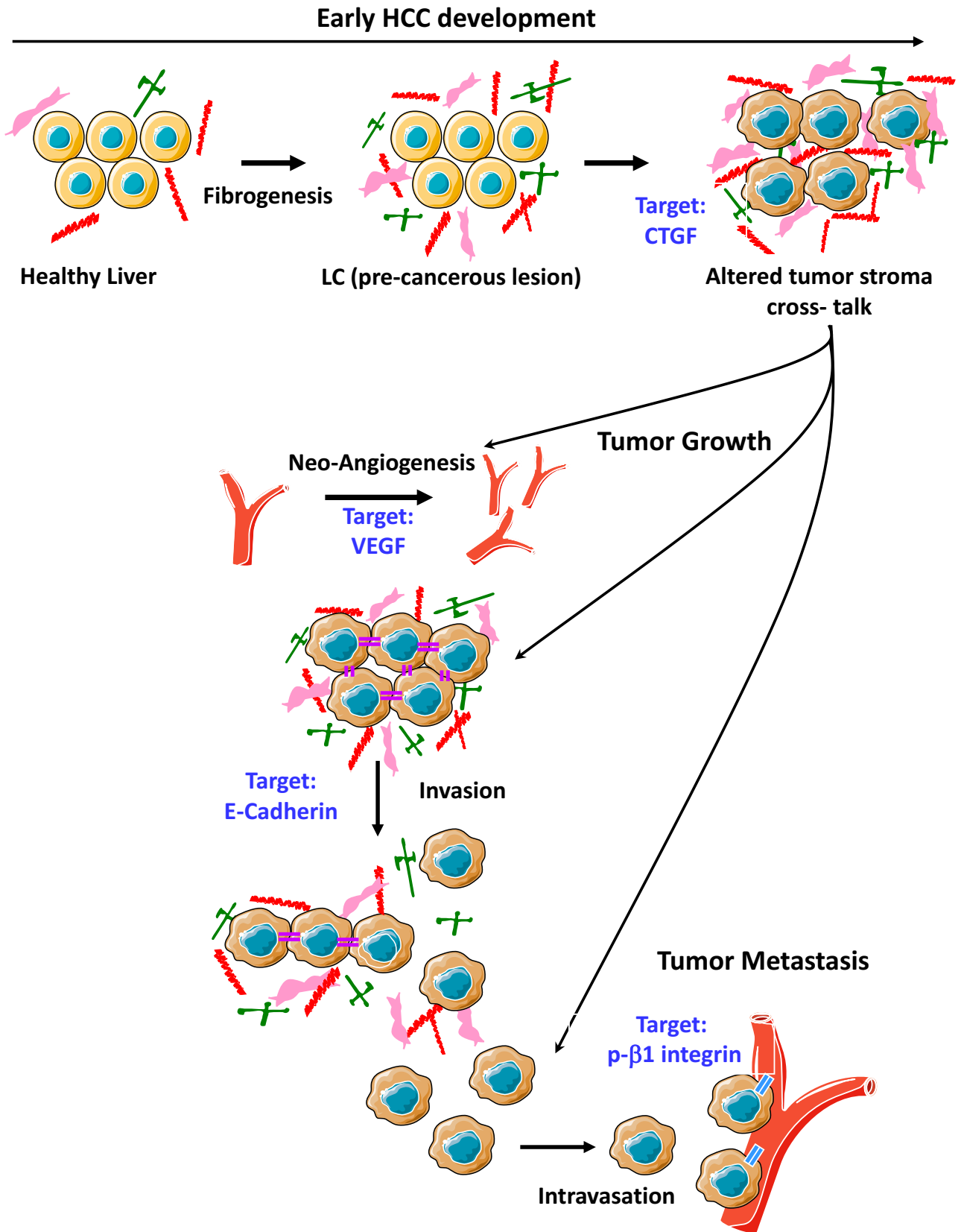
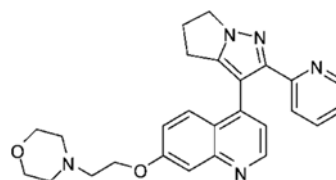


Fig. (1). Schematic representation of the multifunctional contribution of TGF-β1 in HCC progression. TGF-β1 intervenes in various biological steps during HCC progression. It is involved in regulation of the tumor–stroma cross-talk, neo-angiogenesis, intravasation and cancer dissemination. Because of its involvement in multiple steps during tumor progression, blocking the TGF-β1 signaling axis with TGFβR inhibitors could prove to be a promising pharmacological approach to target different steps of HCC progression.

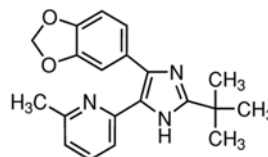
paradigm that by “*hitting one target leads to a domino effect*”. This is easy to understand since the inhibition of a pleiotropic factor like TGF- β 1 indisputably leads to the inhibition of several downstream effectors. The concept of a pharmacological inhibition of TGF- β 1 is different from that of a multikinase inhibitor. The former is based on the fact that blockade of one single target leads to the inhibition of multiple effectors, while the latter involves the inhibition of different kinases to elicit a similar effect [71]. The advantage of “*the domino effect*” is that it has a more efficient mechanism of action, better pharmacodynamics and a lower toxicity. LY2109761 is a selective inhibitor of the TGF- β receptor kinase belonging to a new class of compounds now used to inhibit TGF β R. The chemical structure of LY2109761 and other compounds belonging to different classes of TGF- β receptor kinase inhibitors are shown in (Fig. 2). LY2109761 was discovered during screening to test the structure-activity relationships (SAR) of the quinoline domain of dihydropyrrolopyrazoles [72]. This drug selectively blocks the activation of SMAD2, a marker of TGF β R activation, at concentrations ranging from 0.001 to 0.1 microMolars. Moreover, LY2109761 inhibits migration of HCC cells on fibronectin, laminin-5, and vitronectin, as well as tumor cell invasion through Matrigel at concentrations up to 0.1 microMolar, and up-regulates expression levels of E-cadherin in nonmetastatic HCC tissues treated ex-vivo [73]. LY2109761 has shown an interesting anti-tumor activity against both solid and haematological malignancies. For example, It suppresses pancreatic cancer metastasis to the liver and, in combination with gentacytabin, reduces the tumor mass and increases the survival rate of treated animals [74]. Another study, conducted by Xu *et al.*, showed that LY2109761 inhibits TGF- β 1 produced by bone marrow stromal cells that sustain the survival and chemoresistance of leukaemia cells [75]. Recent studies have shown that LY2109761 decreases metastases to the liver and prolongs survival of mice in a *murine syngeneic model of colon cancer* [76,77]. Another study investigating the capacity of HCC cells to invade the blood vessels, that is one of the major negative prognostic factors in patients with HCC, has shown the impact of inhibition of TGF β R by LY2109761 on this process [78]. In particular, this study showed that invasion of blood vessels is mediated by TGF- β 1, that transactivates β 1 integrin via SMAD2 and SMAD3. TGF- β 1 induces the phosphorylation of the intracytoplasmic tail of β 1 integrin on threonine 788-789 through an “inside-out” mechanism of activation. Because the inhibition of TGF β R by LY2109761 blocks vascular invasion of HCC cells, this may suggest a *rationale* for targeting the TGF- β 1 signaling pathway to inhibit the metastatic dissemination of cancer cells. The antitumor activity of LY2109761 is also associated with the inhibition of molecular pathways involved in neo-angiogenesis and tumor growth of HCC [79]. LY2109761 has shown a promising anti-angiogenic effect that seems to be more effective than the one exerted by bevacizumab, a well-established anti-vascular endothelial growth factor (VEGF) monoclonal antibody. LY210976 blocks the paracrine cross-talk between HCC and endothelial cells and hence the formation of new blood vessels [79]. This effect is mediated by the SMAD2/3 signaling pathway which regulates the secretion of VEGF. Interestingly, LY2109761 has not shown any significant effect on physiological angiogenic development. This study supports the idea that targeting TGF- β signaling in patients with HCC could eventually affect neo-angiogenesis. The take-home message from all these studies is that pharmacological targeting of TGF- β signaling pathway may result in simultaneously blocking several downstream effectors, cellular functions and different processes underlying tumor progression. Blocking one single target, such as TGF β R, which in turn results in inhibition of several downstream functions, is a novel concept. *Hitting one single target that leads to a domino effect* is an attractive and alternative pharmacological concept, but it is not mutually exclusive to that of multi-target/kinase inhibition, in which sorafenib is a paradigm in the treatment of HCC. Therefore, it is plausible that a combination of

Dihydropyrrolopyrazoles



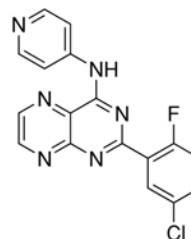
LY2109761

Imidazole-based compounds



SB-505124

Aminopteridine-derivatives



SD-208

Fig. (2). Main classes of TGF- β signaling inhibitors. Some of the recently developed small-molecule inhibitors of the TGF β R are shown. TGF β R inhibitors are based on different scaffolds. LY2109761 (Lilly Research Laboratories) is a dihydropyrrolopyrazole-based compound; SB-505124 (GlaxoSmith Kline) is based on an imidazole scaffold, whereas SD-208 (Scios Inc.) is an aminopteridine-derivative compound.

drugs acting with different mechanisms (i.e. TGF β R inhibitors in association with multikinase inhibitors) may be a promising strategy for the treatment of HCC in the near future.

6. FUTURE PERSPECTIVES

Medical therapy of HCC is an intriguing but not an easy issue, since the pharmacological approach has to take into account the liver cirrhosis that so often underlies human HCC. So far, sorafenib is the sole pharmacological therapy that has revealed some efficiency in terms of improved survival in HCC patients. Sorafenib is an oral multikinase inhibitor that acts on several signaling pathway including the Raf/MEK/ERK pathway, VEGFR and PDGFR [80]. Sorafenib is currently the gold standard therapy to which new anti-HCC drugs must be compared. In addition, new target-based therapy needs to be tailored to the individual patient and therefore the molecular target has to be tested before starting the treatment. This means that if the molecular target is TGF β R, this must be expressed and validated in terms of the specific biological activity in the patient who is a candidate to receive TGF β R inhibitors. This is currently the main challenge that LY2109761 or other TGF β R inhibitors have to overcome to successfully enter the arena of human therapeutic approaches. The availability of a reliable and selective inhibitor is not enough to treat cancer in patients if detailed information on the expression and function of the target is lacking. As

described in this review article, preclinical studies of TGF β R inhibitors (i.e. LY2109761) have recently been conducted. However, more accurate investigations on the drug mechanisms, pharmacokinetics and pharmacodynamics are needed to better evaluate whether this class of inhibitors can rapidly enter early clinical development in HCC.

7. CONCLUDING REMARKS

Despite the dual role of TGF- β in cancer, the use of TGF β R inhibitors is proving to be a promising anti-cancer approach in pre-clinical studies. However, additional studies are needed to confirm the efficacy of this class of inhibitors and to evaluate their toxicity. In HCC, the powerful *rationale* for evaluating TGF- β 1 lies in the fact that TGF- β is involved in tumor progression and regulates several steps related to the interactions with the microenvironment, including the intravasation of tumor cells, angiogenesis and the production of extracellular matrix. In addition, TGF- β promotes liver fibrosis and its inhibition could turn out to be beneficial for HCC patients with underlying liver cirrhosis. While the scientific debate as to whether TGF- β acts as a promoter or suppressor of oncogenesis is still ongoing, a growing body of evidence suggests that inhibition of TGF- β eventually exerts anti-tumor activities. However, these findings must be treated with some caution, and the preliminary data we have obtained so far on TGF- β inhibitors need to be validated in a broader number of studies and in different pre-clinical cancer models before setting up clinical trials. Finally, the ongoing development of new classes of TGF- β inhibitors should improve our arsenal of anti-cancer drugs, as well as promoting a better understanding of the biology of TGF- β in the cancer process.

COMPETING INTERESTS

The authors declare that they have no competing interests.

ACKNOWLEDGMENTS

The Authors are grateful to Mary V. Pragnell, B.A., for language revision.

ABBREVIATIONS

HCC	=	Hepatocellular Carcinoma
TGF- β 1	=	Transforming growth factor-beta
TGF β R	=	Transforming growth factor-beta receptor
AFP	=	Alpha-fetoprotein
CTGF	=	Connective tissue growth factor
CAF	=	Cancer associated fibroblasts
SAR	=	Structure-activity relationships
SMAD2/3	=	Small mother against decapentaplegic
VEGF	=	Vascular endothelial growth factor
MEK	=	Mitogen-activated protein kinase kinase
ERK	=	Extracellular signal-regulated kinase
PDGFR	=	Platelet-derived growth factor

REFERENCES

- Oft M, Heider KH, Beug H. TGFbeta signaling is necessary for carcinoma cell invasiveness and metastasis *Curr Biol* 1998; 8: 1243-52
- Bissell DM. Chronic liver injury, TGF-beta, and cancer *Exp Mol Med* 2001; 33: 179-90
- Llovet JM, Ricci S, Mazzaferro V, et al. Sorafenib in advanced hepatocellular carcinoma *N Engl J Med* 2008; 359: 378-90
- Antonaci S, Giannelli G. Rationale for new drugs targeting the tissue microenvironment in patients with HCC *Curr Pharm Des* 2007; 13: 3288-91
- Roberts LR, Gores GJ. Hepatocellular carcinoma: molecular pathways and new therapeutic targets *Semin. Liver Dis* 2005; 25: 212-25
- Heindryckx F, Colle I, Van VH. Experimental mouse models for hepatocellular carcinoma research *Int J Exp Pathol* 2009; 90: 367-86
- Sanchez A, Fabregat I. Genetically modified animal models recapitulating molecular events altered in human hepatocarcinogenesis *Clin Transl Oncol* 2009; 11: 208-14
- El-Serag HB, Rudolph KL. Hepatocellular carcinoma: epidemiology and molecular carcinogenesis *Gastroenterology* 2007; 132: 2557-76
- Friedman SL, Bissell DM. Hepatic fibrosis: new insights into pathogenesis *Hosp.Pract (Off Ed)* 1990; 25: 43-50
- Bissell DM. Hepatic fibrosis as wound repair: a progress report *J Gastroenterol* 1998; 33: 295-302
- Llovet JM, Di Bisceglie AM, Bruix J, et al. Design and endpoints of clinical trials in hepatocellular carcinoma *J Natl Cancer Inst* 2008; 100: 698-711
- Di MM, Daniele B, Gallo C, Perrone F. Re: Design and endpoints of clinical trials in hepatocellular carcinoma *J Natl Cancer Inst* 2008; 100: 1557-8
- Fausto N, Mead JE, Gruppuso PA, Castilla A, Jakowlew SB. Effects of TGF-beta s in the liver: cell proliferation and fibrogenesis *Ciba Found. Symp* 1991; 157: 165-74
- Thiery JP. Epithelial-mesenchymal transitions in tumour progression. *Nat Rev Cancer* 2002; 2: 442-54
- Massague J. TGFbeta in Cancer *Cell* 2008; 134: 215-30
- Giannelli G, Bergamini C, Fransvea E, Sgarra C, Antonaci S. Laminin-5 with transforming growth factor-beta1 induces epithelial to mesenchymal transition in hepatocellular carcinoma *Gastroenterology* 2005; 129: 1375-83
- Yang L, Moses HL. Transforming growth factor beta: tumor suppressor or promoter? Are host immune cells the answer? *Cancer Res* 2008; 68: 9107-11
- Pardali K, Moustakas A. Actions of TGF-beta as tumor suppressor and pro-metastatic factor in human cancer *Biochim Biophys Acta* 2007; 1775: 21-62
- Bierie B, Moses HL. Tumour microenvironment: TGFbeta: the molecular Jekyll and Hyde of cancer *Nat Rev Cancer* 2006; 6: 506-20
- Diener KR, Need EF, Buchanan G, Hayball JD. TGF-beta signalling and immunity in prostate tumorigenesis *Expert Opin Ther Targets* 2010; 14: 179-92
- Romano MF. Targeting TGFbeta-mediated processes in cancer *Curr Opin Drug Discov Devel* 2009; 12: 253-63
- Cruz-Merino L, Henao-Carrasco F, Garcia-Manrique T, Fernandez-Salguero PM, Codes-Manuel d V. Role of transforming growth factor beta in cancer microenvironment *Clin Transl Oncol* 2009; 11: 715-20
- Gressner OA, Gressner AM. Connective tissue growth factor: a fibrogenic master switch in fibrotic liver diseases *Liver Int* 2008; 28: 1065-79
- Chen XM, Qi W, Pollock CA. CTGF and chronic kidney fibrosis *Front Biosci (Schol.Ed)* 2009; 1: 132-41
- Chu CY, Chang CC, Prakash E, Kuo ML. Connective tissue growth factor (CTGF) and cancer progression *J Biomed Sci* 2008; 15: 675-85
- Mazzocca A, Fransvea E, Dituri F, Lupo L, Antonaci S, Giannelli G. Down-regulation of connective tissue growth factor by inhibition of transforming growth factor beta blocks the tumor-stroma cross-talk and tumor progression in hepatocellular carcinoma *Hepatology* 2010; 51: 523-34
- Gonda TA, Varro A, Wang TC, Tycko B. Molecular biology of cancer-associated fibroblasts: can these cells be targeted in anti-cancer therapy? *Semin.Cell Dev Biol* 2010; 21: 2-10
- Ostman A, Augsten M. Cancer-associated fibroblasts and tumor growth--bystanders turning into key players *Curr Opin Genet Dev* 2009; 19: 67-73
- Mazzocca A, Dituri F, Lupo L, Quaranta M, Antonaci S, Giannelli G. Tumor-secreted lysophosphatidic acid accelerates hepatocellular carcinoma progression by promoting differentiation of peritumoral fibroblasts in myofibroblasts *Hepatology* 2011.
- Muerkoster S, Wegehenkel K, Arlt A, et al. Tumor stroma interactions induce chemoresistance in pancreatic ductal carcinoma cells involving increased secretion and paracrine effects of nitric oxide and interleukin-1beta *Cancer Res* 2004; 64: 1331-7

- [31] Hwang RF, Moore T, Arumugam T, *et al.* Cancer-associated stromal fibroblasts promote pancreatic tumor progression *Cancer Res* 2008; 68: 918-26
- [32] Giannelli G, Mazzocca A, Fransvea E, Lahn M, Antonaci S. Inhibiting TGF-beta signaling in hepatocellular carcinoma *Biochim.Biophys.Acta* 2011; 1815: 214-23
- [33] Roberts AB, McCune BK, Sporn MB. TGF-beta: regulation of extracellular matrix *Kidney Int* 1992; 41: 557-9
- [34] Czaja MJ, Weiner FR, Flanders KC, *et al.* *In vitro* and *in vivo* association of transforming growth factor-beta 1 with hepatic fibrosis. *J Cell Biol* 1989; 108: 2477-82
- [35] Calabrese F, Valente M, Giacometti C, *et al.* Parenchymal transforming growth factor beta-1: its type II receptor and Smad signaling pathway correlate with inflammation and fibrosis in chronic liver disease of viral etiology *J Gastroenterol Hepatol* 2003; 18: 1302-8
- [36] Taniguchi H, Kato N, Otsuka M, *et al.* Hepatitis C virus core protein upregulates transforming growth factor-beta 1 transcription *J Med Virol* 2004; 72: 52-9
- [37] Castilla A, Prieto J, Fausto N. Transforming growth factors beta 1 and alpha in chronic liver disease. Effects of interferon alpha therapy. *N Engl J Med* 1991; 324: 933-40
- [38] Pinzani M, Failli P, Ruocco C, *et al.* Fat-storing cells as liver-specific pericytes. Spatial dynamics of agonist-stimulated intracellular calcium transients. *J Clin Invest* 1992; 90: 642-6
- [39] Yang C, Zeisberg M, Mosterman B, *et al.* Liver fibrosis: insights into migration of hepatic stellate cells in response to extracellular matrix and growth factors *Gastroenterology* 2003; 124: 147-59
- [40] Kawada N. The hepatic perisinusoidal stellate cell *Histol. Histopathol* 1997; 12: 1069-80
- [41] Friedman SL, Roll FJ, Boyles J, Bissell DM. Hepatic lipocytes: the principal collagen-producing cells of normal rat liver *Proc Natl Acad Sci USA* 1985; 82: 8681-5
- [42] George J, Roulot D, Kotliansky VE, Bissell DM. *In vivo* inhibition of rat stellate cell activation by soluble transforming growth factor beta type II receptor: a potential new therapy for hepatic fibrosis *Proc Natl Acad Sci USA* 1999; 96: 12719-24
- [43] Jiang G, Yang HR, Wang L, Wildey GM, Fung J, Qian S, Lu L. Hepatic stellate cells preferentially expand allogeneic CD4+ CD25+ FoxP3+ regulatory T cells in an IL-2-dependent manner *Transplantation* 2008; 86: 1492-502
- [44] Luo JH, Ren B, Keryanov S, *et al.* Transcriptomic and genomic analysis of human hepatocellular carcinomas and hepatoblastomas *Hepatology* 2006; 44: 1012-24
- [45] Winau F, Quack C, Darmono A, Kaufmann SH. Starring stellate cells in liver immunology *Curr Opin Immunol* 2008; 20: 68-74
- [46] Chang KM. Regulatory T cells in hepatitis C virus infection *Hepatol. Res* 2007; 37 Suppl 3: S327-30
- [47] Giannelli G, Antonaci S. Immunological and molecular aspects of liver fibrosis in chronic hepatitis C virus infection *Histol. Histopathol* 2005; 20: 939-44
- [48] Breuhahn K, Longerich T, Schirmacher P. Dysregulation of growth factor signaling in human hepatocellular carcinoma *Oncogene* 2006; 25: 3787-800
- [49] Ito N, Kawata S, Tamura S, *et al.* Elevated levels of transforming growth factor beta messenger RNA and its polypeptide in human hepatocellular carcinoma *Cancer Res* 1991; 51: 4080-3
- [50] Ito N, Kawata S, Tamura S, Shirai Y, Kiso S, Tsushima H, Matsuzawa Y. Positive correlation of plasma transforming growth factor-beta 1 levels with tumor vascularity in hepatocellular carcinoma *Cancer Lett* 1995; 89: 45-8
- [51] Tsai JF, Chuang LY, Jeng JE, *et al.* Clinical relevance of transforming growth factor-beta 1 in the urine of patients with hepatocellular carcinoma *Medicine (Baltimore)* 1997; 76: 213-26
- [52] Giannelli G, Fransvea E, Marinosci F, *et al.* Transforming growth factor-beta1 triggers hepatocellular carcinoma invasiveness via alpha3beta1 integrin. *Am J Pathol* 2002; 161: 183-93
- [53] Yamaguchi R, Yano H, Iemura A, Ogasawara S, Haramaki M, Kojiro M. Expression of vascular endothelial growth factor in human hepatocellular carcinoma. *Hepatology* 1998; 28: 68-77
- [54] Poon RT, Lau CP, Cheung ST, Yu WC, Fan ST. Quantitative correlation of serum levels and tumor expression of vascular endothelial growth factor in patients with hepatocellular carcinoma *Cancer Res* 2003; 63: 3121-6
- [55] Jonas S, Bechstein WO, Steinmuller T, *et al.* Vascular invasion and histopathologic grading determine outcome after liver transplantation for hepatocellular carcinoma in cirrhosis *Hepatology* 2001; 33: 1080-6
- [56] Giannelli G, Pierri F, Trerotoli P, *et al.* Occurrence of portal vein tumor thrombus in hepatocellular carcinoma affects prognosis and survival. A retrospective clinical study of 150 cases *Hepatol Res* 2002; 24: 50-9
- [57] Sumie S, Kuromatsu R, Okuda K, *et al.* Microvascular invasion in patients with hepatocellular carcinoma and its predictable clinicopathological factors. *Ann Surg Oncol* 2008; 15: 1375-82
- [58] Sakata J, Shirai Y, Wakai T, Kaneko K, Nagahashi M, Hatakeyama K. Preoperative predictors of vascular invasion in hepatocellular carcinoma. *Eur J Surg Oncol* 2008.
- [59] Bedossa P, Peltier E, Terris B, Franco D, Poynard T. Transforming growth factor-beta 1 (TGF-beta 1) and TGF-beta 1 receptors in normal, cirrhotic, and neoplastic human livers. *Hepatology* 1995; 21: 760-6
- [60] Tsai JF, Jeng JE, Chuang LY, *et al.* Clinical evaluation of urinary transforming growth factor-beta1 and serum alpha-fetoprotein as tumour markers of hepatocellular carcinoma *Br J Cancer* 1997; 75: 1460-6
- [61] Song BC, Chung YH, Kim JA, *et al.* Transforming growth factor-beta1 as a useful serologic marker of small hepatocellular carcinoma *Cancer* 2002; 94: 175-80
- [62] Murawaki Y, Ikuta Y, Nishimura Y, Koda M, Kawasaki H. Serum markers for fibrosis and plasma transforming growth factor-beta 1 in patients with hepatocellular carcinoma in comparison with patients with liver cirrhosis *J Gastroenterol Hepatol* 1996; 11: 443-50
- [63] Abreu JG, Ketpura NI, Reversade B, De Robertis EM. Connective-tissue growth factor (CTGF) modulates cell signalling by BMP and TGF-beta. *Nat Cell Biol* 2002; 4: 599-604
- [64] Bennewith KL, Huang X, Ham CM, *et al.* The role of tumor cell-derived connective tissue growth factor (CTGF/CCN2) in pancreatic tumor growth. *Cancer Res* 2009; 69: 775-84
- [65] Leask A, Abraham DJ. All in the CCN family: essential matricellular signaling modulators emerge from the bunker *J Cell Sci* 2006; 119: 4803-10
- [66] Lacher MD, Tiirikainen MI, Saunier EF, *et al.* Transforming growth factor-beta receptor inhibition enhances adenoviral infectability of carcinoma cells via up-regulation of Coxsackie and Adenovirus Receptor in conjunction with reversal of epithelial-mesenchymal transition. *Cancer Res* 2006; 66: 1648-57
- [67] Yingling JM, Blanchard KL, Sawyer JS. Development of TGF-beta signalling inhibitors for cancer therapy. *Nat Rev Drug Discov* 2004; 3: 1011-22
- [68] Arteaga CL. Inhibition of TGFbeta signaling in cancer therapy. *Curr Opin Genet Dev* 2006; 16: 30-7
- [69] Wrzesinski SH, Wan YY, Flavell RA. Transforming growth factor-beta and the immune response: implications for anticancer therapy. *Clin Cancer Res* 2007; 13: 5262-70
- [70] Biswas S, Criswell TL, Wang SE, Arteaga CL. Inhibition of transforming growth factor-beta signaling in human cancer: targeting a tumor suppressor network as a therapeutic strategy. *Clin Cancer Res* 2006; 12: 4142-6
- [71] Fransvea E, Mazzocca A, Santamato A, Azzariti A, Antonaci S, Giannelli G. Kinase activation profile associated with TGF-beta-dependent migration of HCC cells: a preclinical study *Cancer Chemother Pharmacol* 2011; 68: 79-86
- [72] Li HY, McMillen WT, Heap CR, *et al.* Optimization of a dihydropyridopyrazole series of transforming growth factor-beta type I receptor kinase domain inhibitors: discovery of an orally bioavailable transforming growth factor-beta receptor type I inhibitor as antitumor agent. *J Med Chem* 2008; 51: 2302-6
- [73] Fransvea E, Angelotti U, Antonaci S, Giannelli G. Blocking transforming growth factor-beta up-regulates E-cadherin and reduces migration and invasion of hepatocellular carcinoma cells *Hepatology* 2008; 47: 1557-66
- [74] Melisi D, Ishiyama S, Sclabas GM, *et al.* LY2109761, a novel transforming growth factor beta receptor type I and type II dual inhibitor, as a therapeutic approach to suppressing pancreatic cancer metastasis *Mol Cancer Ther* 2008; 7: 829-40
- [75] Xu Y, Tabe Y, Jin L, *et al.* TGF-beta receptor kinase inhibitor LY2109761 reverses the anti-apoptotic effects of TGF-beta1 in myelo-monocytic leukaemic cells co-cultured with stromal cells. *Br J Haematol* 2008; 142: 192-201

- [76] Zhang B, Halder SK, Zhang S, Datta PK. Targeting transforming growth factor-beta signaling in liver metastasis of colon cancer. *Cancer Lett* 2009; 277: 114-20
- [77] Zhang B, Halder SK, Kashikar ND, *et al.* Antimetastatic role of Smad4 signaling in colorectal cancer *Gastroenterology* 2010; 138: 969-80
- [78] Fransvea E, Mazzocca A, Antonaci S, Giannelli G. Targeting transforming growth factor (TGF)-betaRI inhibits activation of beta1 integrin and blocks vascular invasion in hepatocellular carcinoma *Hepatology* 2009; 49: 839-50
- [79] Mazzocca A, Fransvea E, Lavezzari G, Antonaci S, Giannelli G. Inhibition of transforming growth factor beta receptor I kinase blocks hepatocellular carcinoma growth through neo-angiogenesis regulation *Hepatology* 2009; 50: 1140-51
- [80] Abou-Alfa GK, Schwartz L, *et al.* Phase II study of sorafenib in patients with advanced hepatocellular carcinoma. *J Clin Oncol* 2006; 24: 4293-300

Received: March 6, 2012

Accepted: March 18, 2012